



3rd National Conference on Environmental Sampling and Detection for Bio-Threat Agents

Sampling Tools for First Responders

**High Efficiency Bio-Agent Recovery from
Simulated Environmental Surfaces with Robust
Liquid Rinse Vacuum Collection System**

Kevin J. Church

Laboratory Director

Microbial-Vac Systems, Inc.



Sampling Tools for First Responders

- In response to a biological incident/attack, First Responders must have sampling devices with the capabilities to efficiently and safely collect samples.
 1. Sampling is the initial step in determining the nature of the contamination
 2. There is a need for new sampling tools and technology
- The ultimate goal of any sampling device/method is to provide a representative sample of the bio-burden on a surface of interest and successfully deliver that sample to a detection device.
- This presentation will focus on a new sampling device: The M-Vac



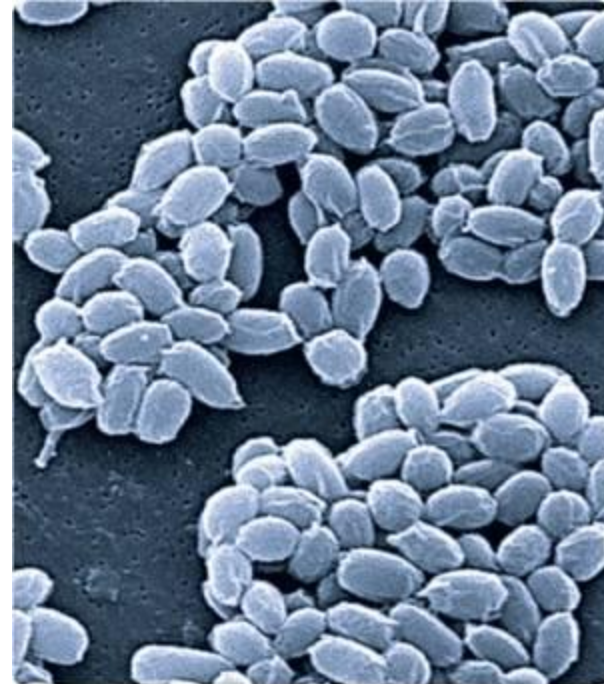
Sampling Challenges for First Responders

- Bio-agents may be widely dispersed over a large area
- Samples must be taken off a variety of surfaces with a high degree of confidence
- Surface levels may be very low
- A number of potential bio-agents can cause disease at extremely low levels
 1. Q-Fever: 1 organism can cause disease in a susceptible person
 2. Tularemia: Approximately 10-50 cells can cause disease
 3. Anthrax: Estimated $\approx 10,000$ cells. May be significantly less for susceptible people



List of High Priority Threat Bio-Agents:

- Anthrax
- *Burkholderia* bacteria
- *C. botulinum* toxins
- Plague
- Q-fever
- Smallpox
- Typhus
- Tularemia
- Various viral hemorrhagic fevers



Bacillus anthracis spores



Sampling Challenges: Determining Sampling Efficiency

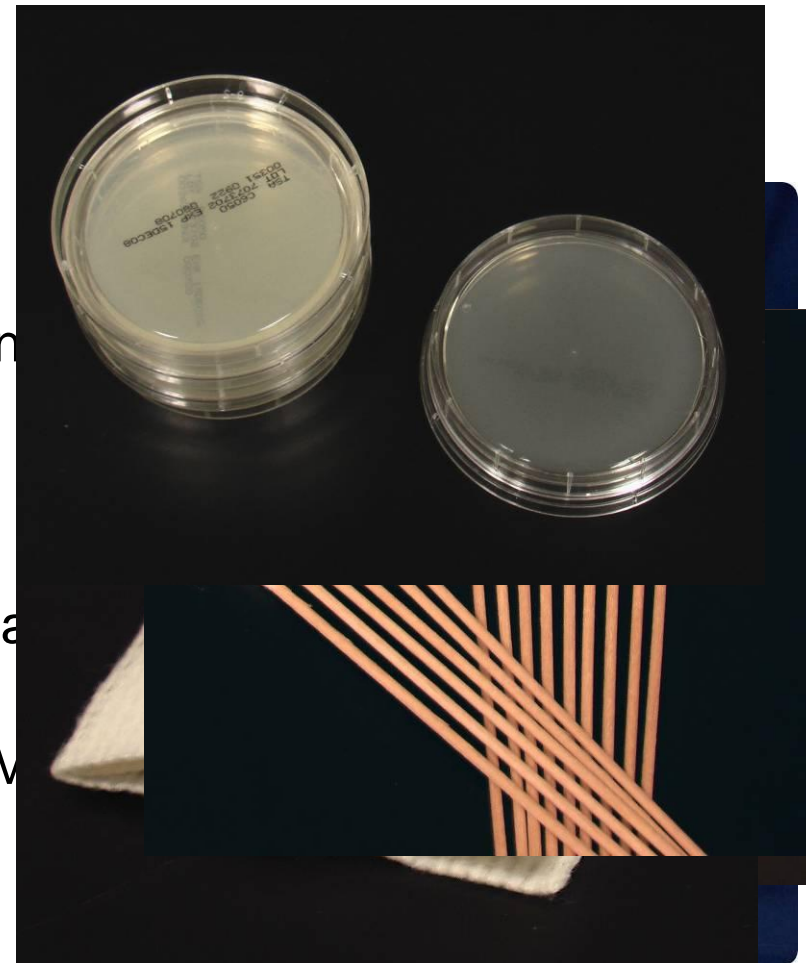
Sampling efficiency: A measurement of a sampling device's ability to effectively sample a surface of interest. Involves 3 main steps:

1. Location/discovery: Locating target organisms on the surface of interest.
2. Extraction: Extracting the target organism from the surface of interest with the sampling device.
3. Elution: Removing the target organism from the sampling device into solution for detection.



Currently Utilized Sampling Devices

1. Swabs: cotton, rayon, polyester,
2. Sponges: cellulose based
3. Surface Wipes: commonly cotton
4. HEPA Vacuum Socks
5. Agar Contact: contact plates, agar
6. The Microbial-Vacuum System (MVS)





The M-Vac System



SEC with attached MS-Kit

- Non-destructive wet-vacuum surface sampling device
- Applies a sterile surface rinse solution (SRS) at a constant pressure
- Vacuum pressure capture of surface suspension at 15 in. Hg (-50 kPa)
- Collects up to 150 ml of surface suspension per sample bottle. Delivers 160 ml/min.
- Unique filter system ensures that the sample is isolated to the collection bottle.

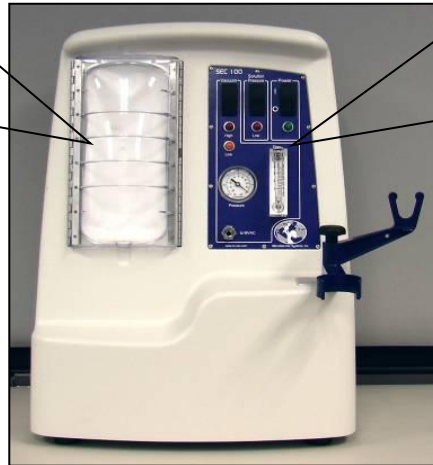


The M-Vac System Overview



Solution Pressurization Chamber

Support Equipment Case (SEC)



SEC Control Panel

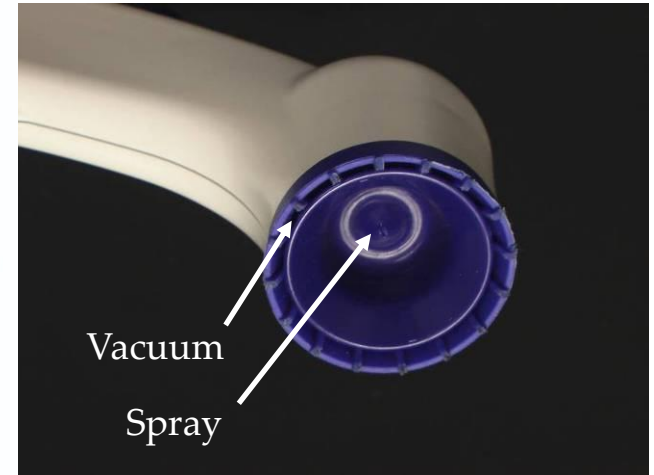
- SEC footprint: 19"x16"x25" (48.3x40.6x63.5 cm), 58 lbs (26.3 kg), portable with "suitcase style" handle and wheels. 110 volts AC, 220 volt system also available



The M-Vac System Overview



SRS Bag



SRS: PBS, Neutralizing Buffer, Butterfield's Buffer, etc.



Sampling Efficiency

1. Location/discovery: Locating target organisms on the surface of interest.
 - a. Location efficiency of sampling device is determined by:
 - i. Concentration of bio-agent (average # bio-agent per surface area)
 - ii. Distribution of bio-agent (uniform vs variable)
 - iii. Surface area of each sample
 - iv. Number of samples

2. **Current Swab/Swipe Devices**
 - a. Small surface area per sample: typically 25 cm² for swabs, 100 cm² for sponges
 - b. Large number of samples needed

3. **M-Vac**
 - a. Large surface area per sample: up to 1600 cm².
 - b. Fewer samples needed

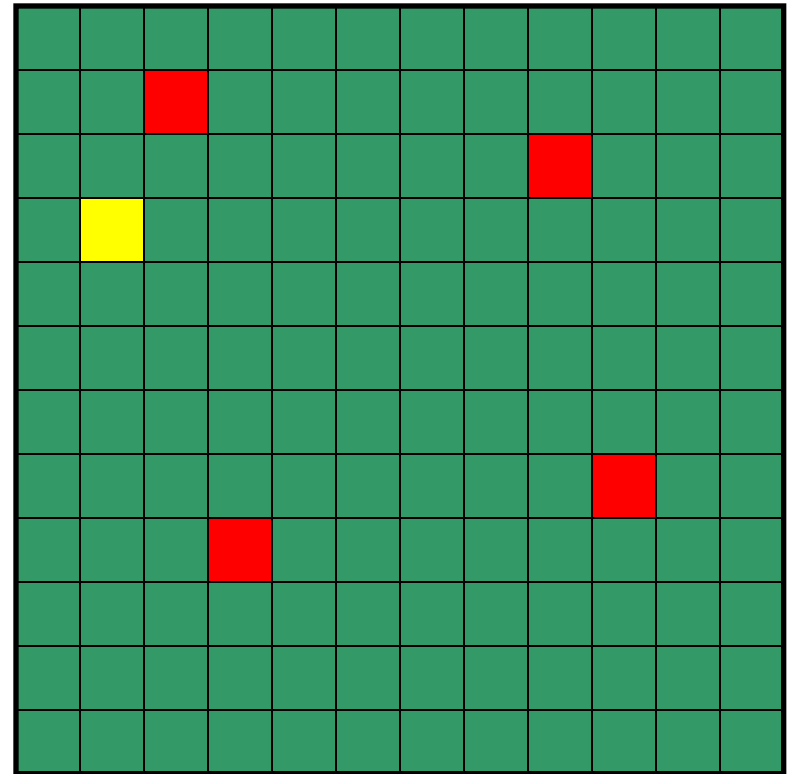


Location Efficiency

 = Contamination

 = Swab sample sites*

*Swab sample sites are 5x5 cm
~25 cm²



**3600 sq cm surface with 5x5 cm grid
and 4 isolated sites of contamination**

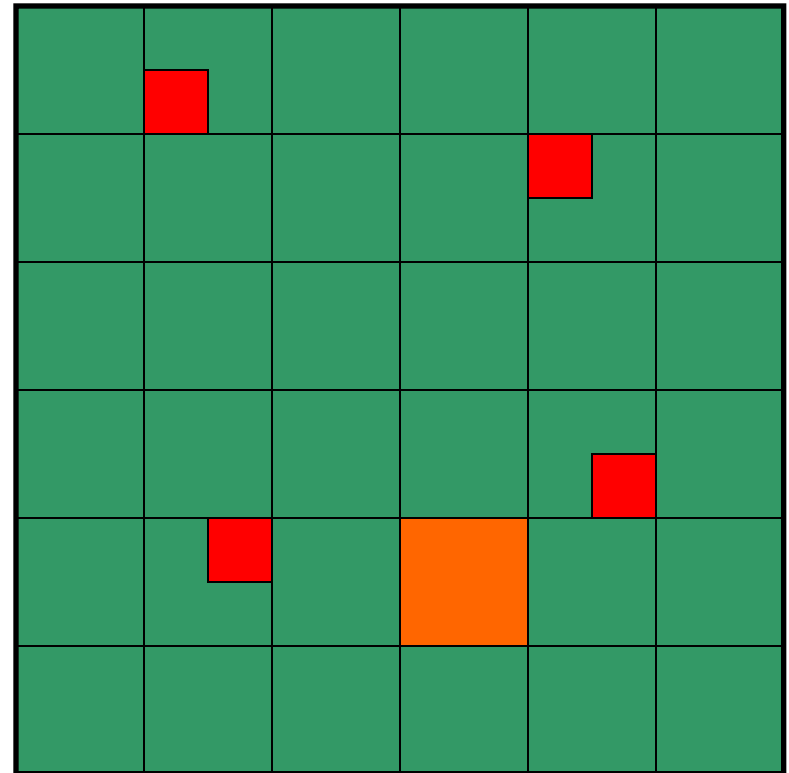


Location Efficiency

 = Contamination

 = Sponge sample sites*

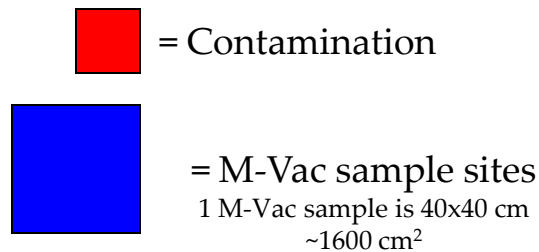
*Sponge sample sites are 10x10 cm
~ 100 sq cm



**3600 sq cm surface with 10x10 cm grid
and 4 isolated sites of contamination**

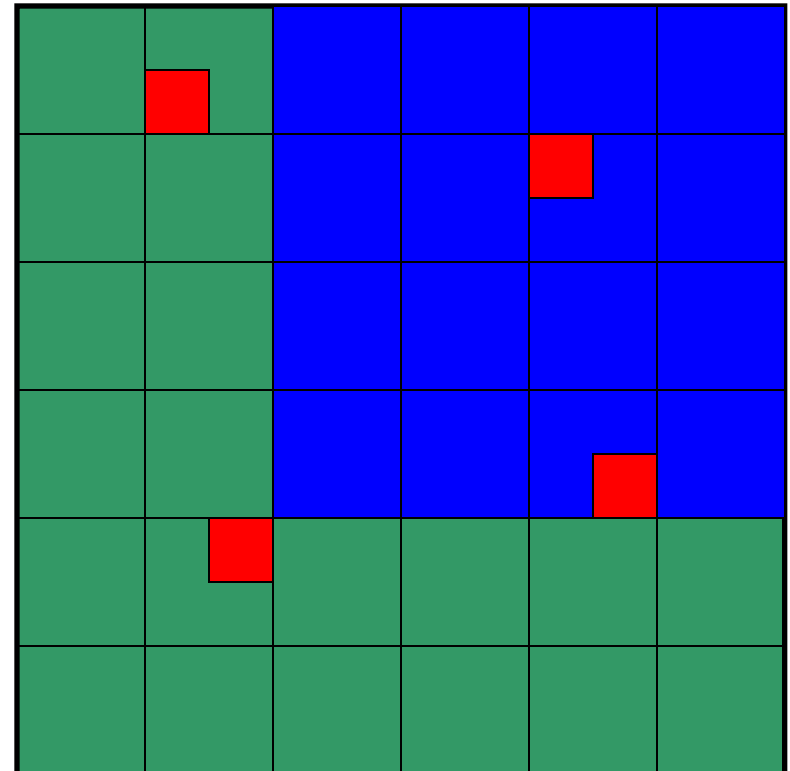


Location Efficiency



Method	# of samples	Total surface area covered (sq cm)	P*
Swab	64	1060	92%
Sponge	16	1600	92%
M-Vac	1	1600	91%

*Probability of finding at least 1 contamination site with a random sample within the given area. Assumes sampling a contaminated site would result in a positive detection result.



3600 sq cm Surface with 10x10 cm grid and 4 isolated sites of contamination



Recovery Efficiency

1. Recovery Efficiency: Number of organisms recovered from a surface by a sampling device from a known population of organisms. Includes extraction and elution steps.

2. Current Swab/Swipe Devices
 - a. Historically low rates of recovery*
 - b. Difficulty in sampling rough or porous surfaces**
 - c. Retain organisms within device**

3. M-Vac
 - a. Experimentally high rates of recovery
 - b. Can sample rough or porous surfaces more efficiently
 - c. No elution step

*Rose L, Jensen B, Peterson A, Banerjee SN, Arduino MJ. Swab materials and *Bacillus anthracis* spore recovery from nonporous surfaces. Emerg Infect Dis [serial on the Internet]. 2004 June [11/14/08]. Available from: <http://www.cdc.gov/ncidod/EID/vol10no6/03-0716.htm>

**Sanderson WT, Hein MJ, Taylor L, Curwin BD, Kinnes GM, Seitz TA, et al. Surface sampling methods for *Bacillus anthracis* spore contamination. Emerg Infect Dis [serial online] 2002 Oct [11/14/08];8. Available from: URL: <http://www.cdc.gov/ncidod/EID/vol8no10/02-0382.htm>



Comparison of Recovery Efficiency

Outline of Experimental Procedures

1. Inoculation with *B. subtilis* spores

- a. Approx. 1.5×10^2 spores per coupon, very low level
- b. Spot inoculate onto sterile environmental surfaces
 - i. Stainless steel, HDPE, ceramic tile, carpet, polypropylene
 - ii. Sampling area of 100 cm² on all coupons
- c. Inoculum spread evenly, allowed to dry

2. Surface Sampling

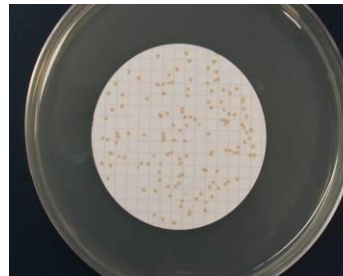
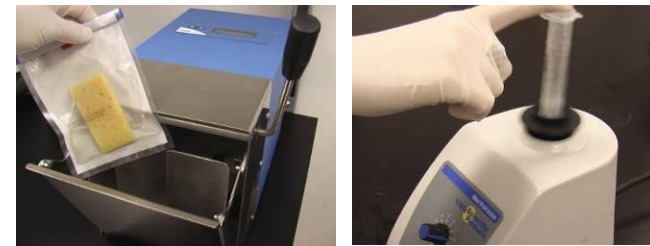
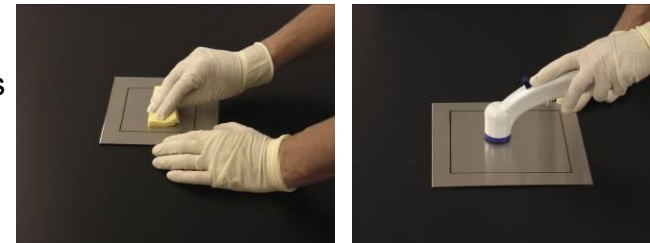
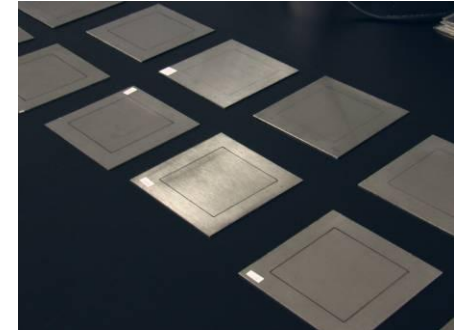
- a. Sample devices were used per manufacturer's instructions

3. Sample Processing

- a. Elution of spores from sampling device
 - i. Stomaching – sponges and gauze
 - ii. Vortexing – swabs

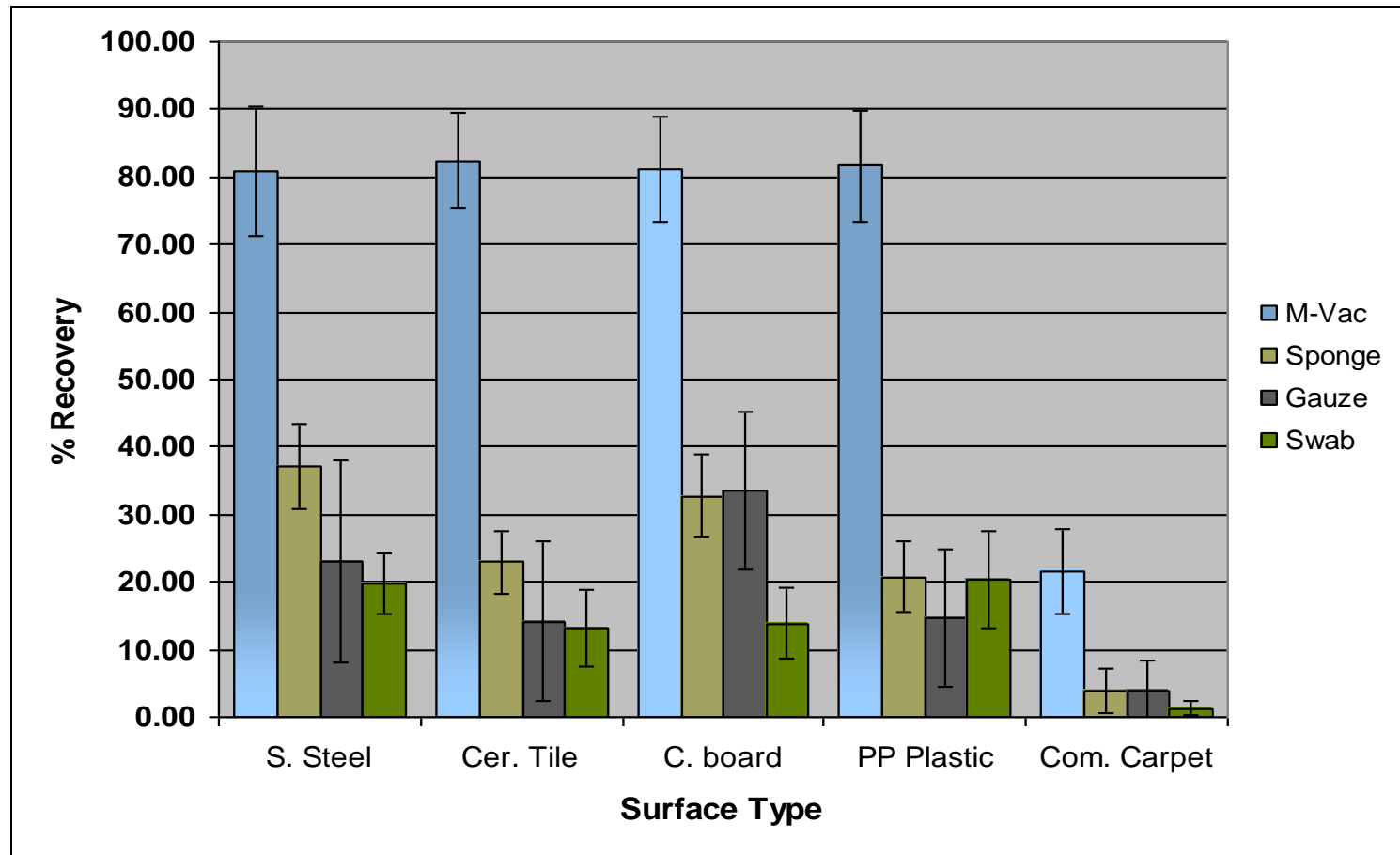
4. Quantitation

- a. Membrane filtration
- b. Counts following 24h incubation





Recovery of *B. subtilis* Spores from Simulated Environmental Surfaces



n=25 for all treatments



Summary

Advantages of the M-Vac

- Larger surface area per sample
- Reduced number of samples
- No elution step
- Higher rate of recovery from a variety of surfaces



Contributors:

**Jared Maughan
Kelly Black
Dr. Bruce Bradley**



Questions?